

Bioactivation of Furans by Human Lung and Liver Microsomes and S9

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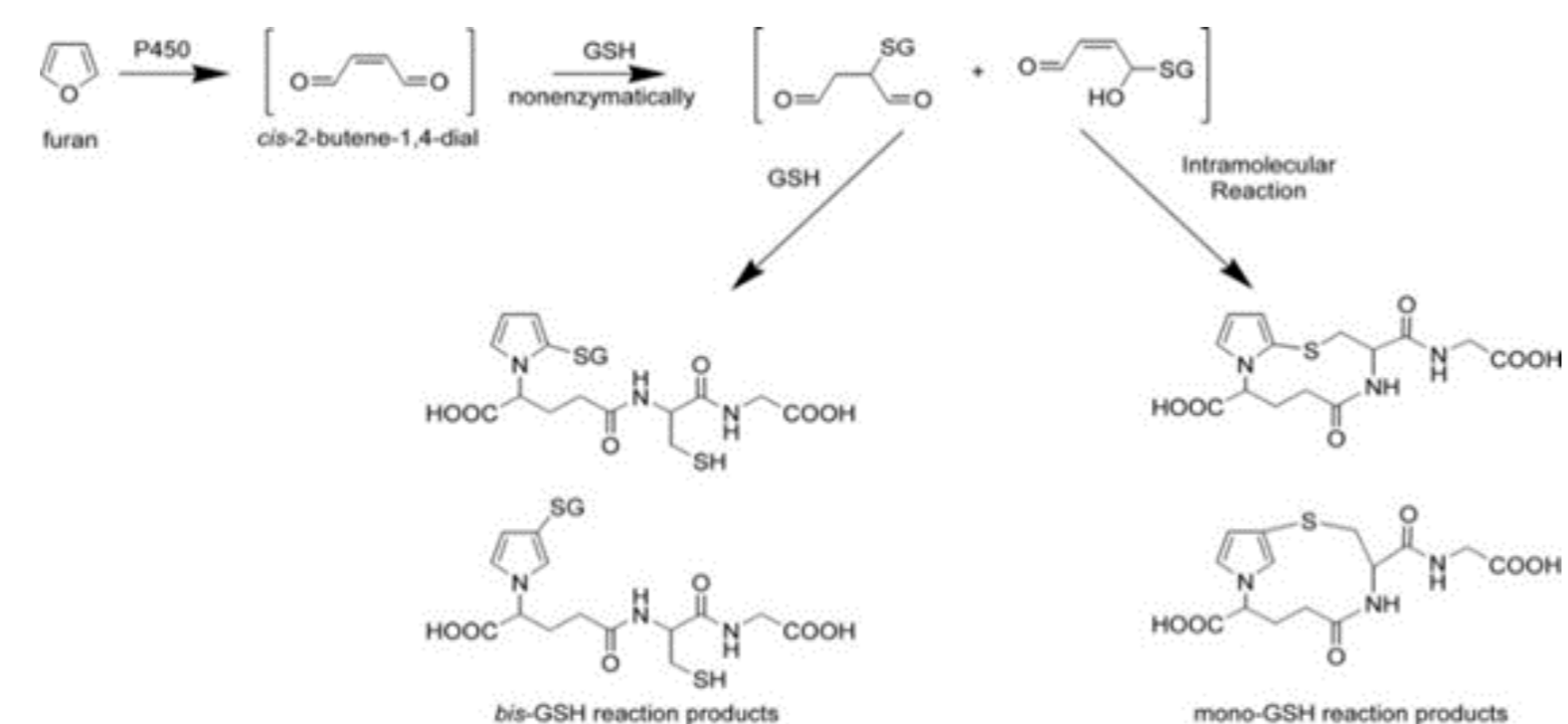
Introduction

Background

Direct exposure to inhaled mainstream cigarette smoke is known to cause damage in the human lung. Although the lung has a significant capability for biotransforming many of the harmful or potentially harmful compounds (HPHCs) present in cigarette smoke, thereby reducing acute toxicity, in some instances biotransformation can result in the generation of more reactive, and thus more toxic, metabolites. This is the case for furan and simple alkylfurans¹.

- Furans are naturally present in tobacco leaves and are also formed by thermal degradation of carbohydrate precursors (between 300-400°C)² during the burning or heating of tobacco
- Biotransformation of simple furans in the body can be described by two major pathways, either by the initial generation of reactive intermediates (cis-2-butene-1,4-dial and derivatives) by CYP2E1 metabolism³ or, if substituted with a side chain (e.g., furfuryl alcohol), by oxidation to the corresponding aldehydes and acids prior to Phase II conjugation and excretion from the body

General scheme for furan bioactivation by CYP2E1 and subsequent conjugation with glutathione



Aim

To evaluate the capability of the human lung (by using subcellular fractions as a surrogate model) to generate reactive intermediate metabolites (RIMs) from the biotransformation of simple furans by Phase I oxidation, and Phase II conjugation with activated sulfate, generating potentially toxic sulfate conjugates, with a view to establish a link between exposure to such compounds and disease-relevant mechanisms (e.g., oxidative stress, inflammation, apoptosis).

To compare the kinetic parameters of human lung (smoker and non-smoker) versus human liver subcellular fractions for the metabolism of simple furans.

Strategy

- Establish a robust and easy to use *in vitro* model to characterize the metabolic fate of furans in human liver and lung subcellular fractions (microsomes as well as S9 for studying Phase I or Phase I+II metabolism)
- Evaluate kinetic parameters for parent compound elimination and metabolite formation for a simple mixture of furans and for cigarette smoke (from 3R4F reference cigarette⁴ bubbled through phosphate buffered saline (wsPBS)) containing furans as part of a complex mixture
- Analyze reactive intermediate metabolites by determination of their glutathione adducts and reactive sulfate conjugates using liquid chromatography coupled with high resolution mass spectrometry (LC-HR-MS)

Materials and Methods

Test/Reference Material and Application Levels

System:	Human Microsomes (Phase I)				Human S9 (Phase I + II)	
	Liver	Lung	Liver	Lung	Liver	Lung
Organ:	Furan-Mix		Ref. Cig. 3R4F		Ref. Cig. 3R4F	
Test Item:	Furan-Mix		Ref. Cig. 3R4F		Ref. Cig. 3R4F	
Smoke Generation:	-		Health Canada (HC)		Health Canada (HC)	
Application level:	1µM	10µM	10µM	10µM	0.08 cig./incub.	0.08 cig./incub.
Protein/incubation:	0.5mg	0.5mg	2mg	2mg	2mg	2mg
Furan	2.8	27.5	6.9	6.8	3.4	3.4
2-Methylfuran	2.2	22.2	5.5	10.3	5.2	5.2
2,5- & 2,4-Dimethylfuran	3.8	37.6	9.4	10.4	5.2	5.2
2-Acetylfuran	2.0	20.0	5.0	2.3	1.2	1.2
Furfural	2.4	24.1	6.0	15.2	7.6	7.6
5-Methylfurfural	2.0	20.1	5.0	4.7	2.4	2.4
2-Furanmethanol	2.3	23.0	5.8	0.6	0.3	0.3
5-Methyl-2-furanmethanol	1.9	19.1	4.8	0.07	0.04	0.04
5-Hydroxymethyl-furfural	1.9	19.1	4.8	15.9	8.0	8.0

Microsomal and S9 Incubations

- Metabolites were generated *in vitro* using an enzymatic system containing human liver microsomes and S9 fractions combined with a NADPH Regenerating System (NRS) containing NADP, G6P and G6PDH as well as alamethicin and co-factors required for Phase II conjugation (e.g. PAPS, UDPGA...)
- Aqueous smoke samples were pre-incubated with glutathione (GSH) for 30 minutes in order to prevent an irreversible inhibition of CYPs by smoke electrophiles (e.g. acrolein, crotonaldehyde...)
- Reactive intermediate metabolites were trapped using GSH as a soft nucleophile (selected experiments used a mixture of stable isotope labeled (glycine-¹³C₂, ¹⁵N) and non-labeled GSH)
- Individual vials for each incubation time point were used to avoid any loss of volatile compounds
- To avoid adsorption effects at material surfaces, silanized glass was used
- Corresponding negative controls followed the same procedure in the absence of the NRS
- Enzymatic activity was stopped by the addition of methanol, containing a mixture of internal standards, followed by crash freezing using dry ice, thawing and centrifugation
- Supernatants were directly injected onto GCxGC-TOF-MS (cool-on-column) and LC-HR-MS systems

LC-HR-MS analysis

LC system:
Thermo Fisher Scientific Accela 1250™ LC
Injection volume: 5 µL
Eluent A: 0.1% formic acid in water
Eluent B : 0.1% formic acid in acetonitrile
Flow rate: 400 µL/min
Column: Thermo Hypersil Gold aQ (150 x 2.1 mm, 1.9 µm)
HPLC gradient:
0-3min (0%B) ->15min (20%B) ->16-19min (100%B)

MS system:
Thermo Fisher Scientific Q Exactive™
Ionization mode: ESI (+) / (-)
Scanning mode: full scan
Scan range: 80 – 800 m/z
Resolution: 70000



GCxGC-TOF-MS analysis

GCxGC system:
Agilent 6890A with ZOE Modulator
Injection: cool-on-column, 0.5µL
1D column: SLB-IL59 (30m x 0.25mm x 0.20µm)
2D column: SLB-IL76 (2m x 0.25mm ID x 0.20µm)
T-program 1D: 30°C (2min) – 5°C/min – 250°C (15min)
T-program 2D: 35°C (2min) – 4°C/min – 43°C – 5.5°C/min – 270°C (15min)
Modulation period: 6sec (1sec hot pulse)

MS system:
LECO Pegasus® 4D
Ionization mode: EI (70eV)
Scan range: 35 – 500 m/z,
Acquisition rate: 200Hz
Resolution: Unit



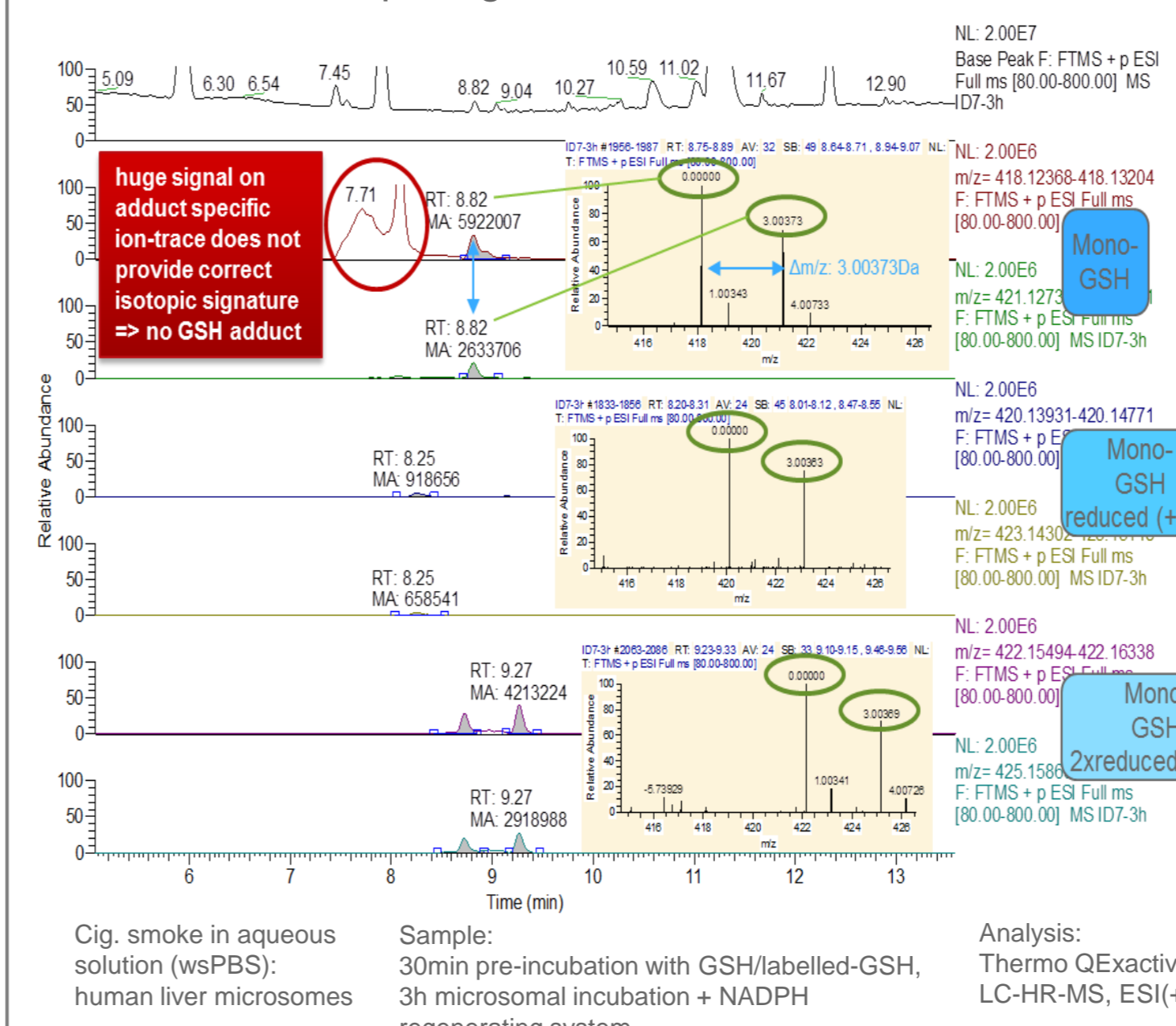
Results

Glutathione Conjugates for Reactive Intermediate Metabolites of Furans by Phase I Metabolism – the Downstream Modification Challenge

Conjugation of RIMs derived from furans with GSH form a multiplicity of initial adducts (1,4-Michael-addition adducts, intramolecular rearrangement products and bis-GSH-adducts), which are prone to subsequent metabolic processes (e.g. reduction, already known for acrolein-glutathione adducts). The presence of downstream reduced GSH-adducts from furans was demonstrated using a mixture of stable isotope labeled and non-labeled GSH as a trapping system. The presence of carbonyl reductase activity within the subcellular fractions could be confirmed by the reduction of 3-oxopropyl-GSH (initial reaction product of acrolein and GSH) to its reduced form, 3-hydroxypropyl-GSH.

Downstream reduction of initial RIM glutathione adducts

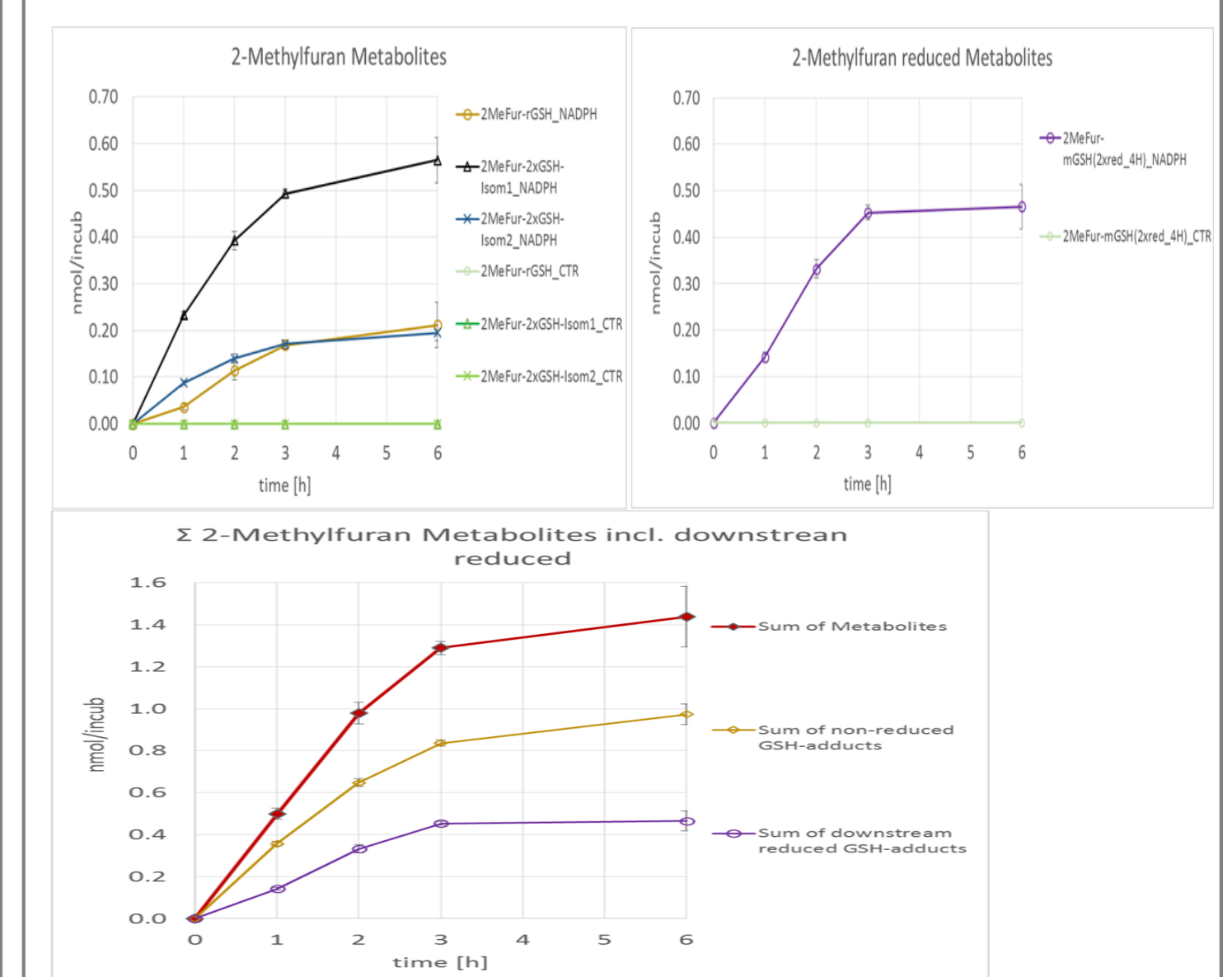
Example: RIMs of 2,5- & 2,4-dimethylfuran conjugated to stable isotope labeled/non-labeled GSH provides a characteristic isotopic signature of $\Delta m/z$: 3.00374Da



Furans RIM glutathione adducts, examples

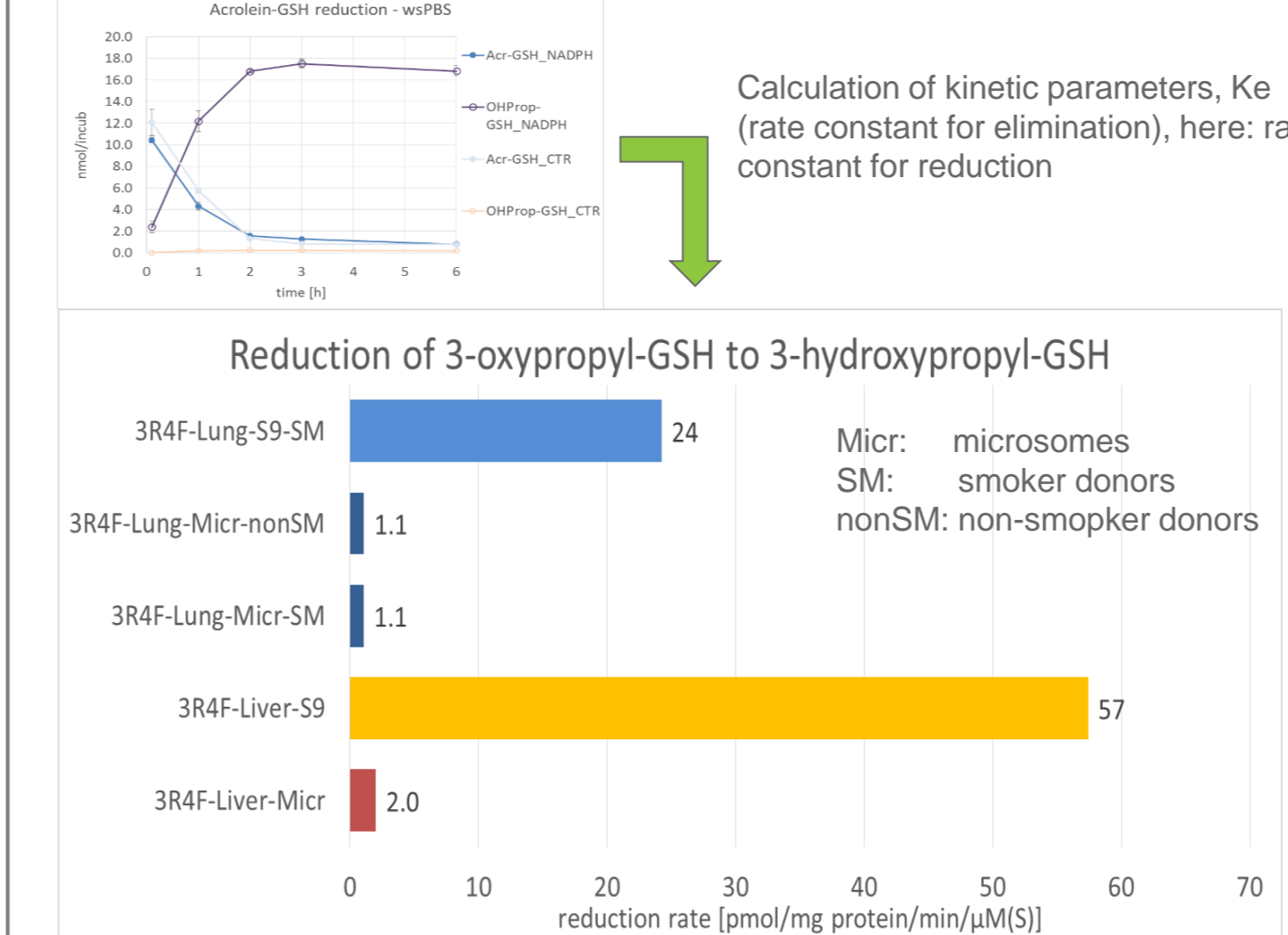
For the evaluation of metabolite formation rates, all currently known types of RIM adducts were considered. Examples below illustrate the diversity of adducts formed.

2-Methylfuran in furan-mix (1µM), liver microsomes, N=3



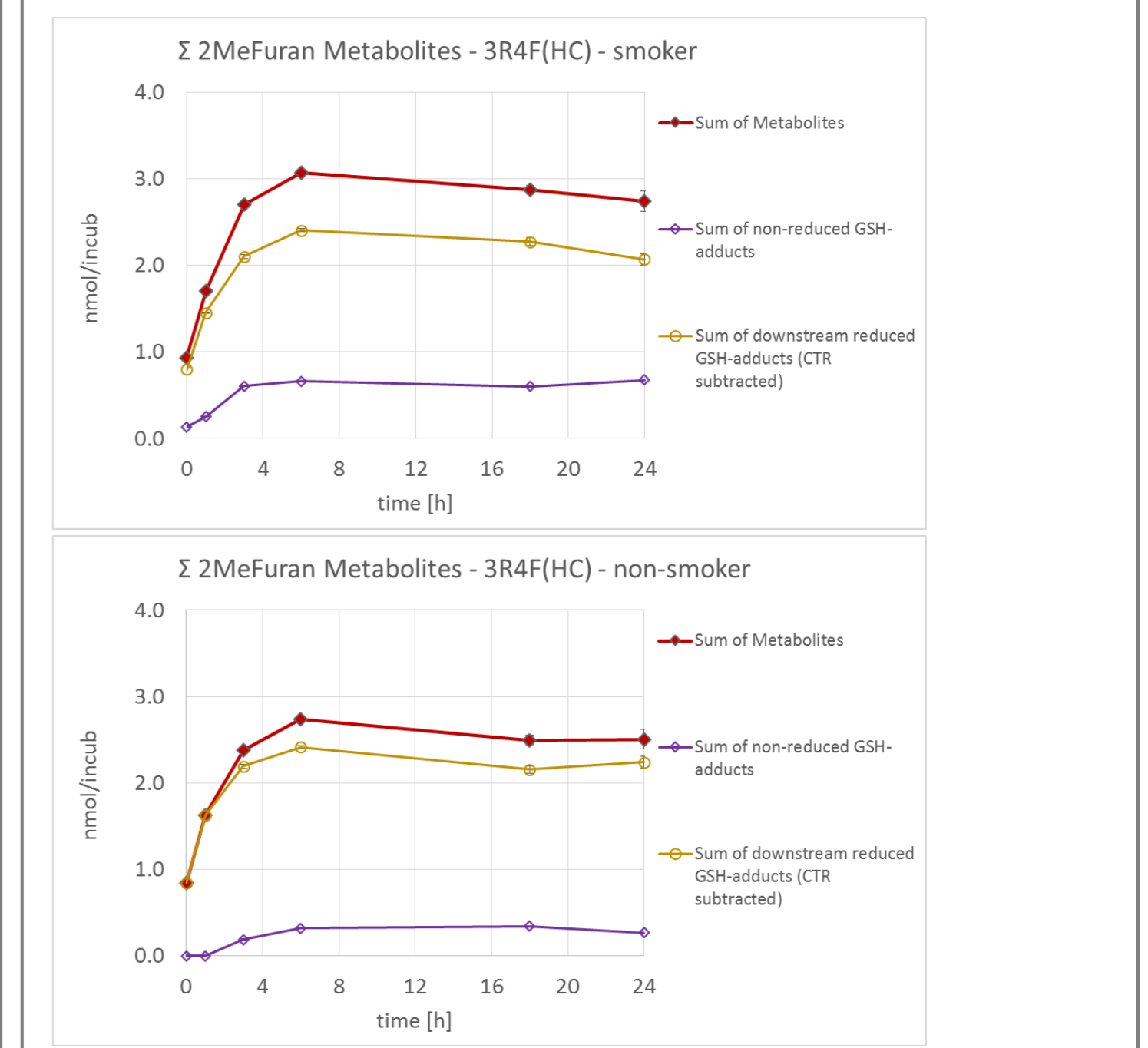
Carbonyl reductase activity for subcellular fractions

Reduction of initial acrolein-GSH adduct: lung microsomes, N=3



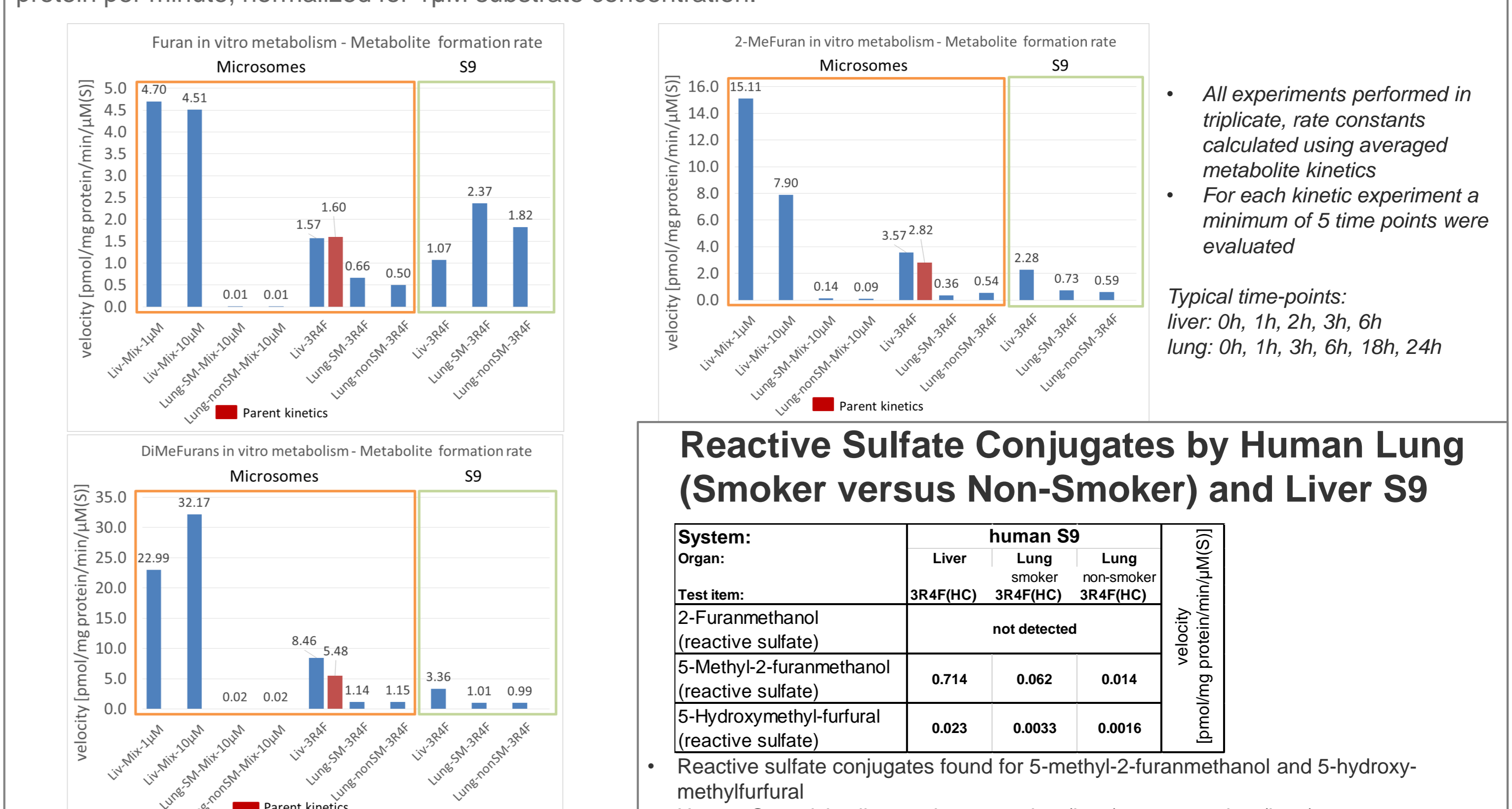
- All tested subcellular fractions showed reductase activity
- Consequently all metabolic experiments included screening for the presence of reduced RIM glutathione adducts

2-Methylfuran - wsPBS-3R4F (6cig./36mL, PBS)



Formation rate of Reactive Intermediate Metabolites by Human Lung (Smoker versus Non-Smoker) and Human Liver Microsomal Subcellular Fractions

The rate of metabolite formation was evaluated using log transformed averaged kinetics data (N=3) for the sum of all glutathione adduct species for each RIM. Values for velocity of metabolite formation are expressed as pmol metabolite formed per mg protein per minute, normalized for 1µM substrate concentration.



Reactive Sulfate Conjugates by Human Lung (Smoker versus Non-Smoker) and Liver S9

System:	human S9		
	Liver	Lung smoker	Lung non-smoker
Organ:	3R4F(HC)	3R4F(HC)	3R4F(HC)
Test Item:	2-Furanmethanol (reactive sulfate)	not detected	not detected
5-Methyl-2-furanmethanol (reactive sulfate)	0.714	0.062	0.014
5-Hydroxymethyl-furfural (reactive sulfate)	0.023	0.0033	0.0016

- Reactive sulfate conjugates found for 5-methyl-2-furanmethanol and 5-hydroxymethylfurfural
- Human S9 activity: liver >> lung, smoker (lung) > non-smoker (lung)

Conclusions

- Human lung subcellular fractions are able to toxify furan and simple alkylfurans by CYP-related Phase I metabolism by generating reactive electrophilic intermediates
- Conjugation of RIMs derived from furans with GSH forms a multiplicity of initial adducts, which are also subject to subsequent metabolic reduction, greatly complicating the evaluation process
- The formation rate for furans reactive metabolites was significantly lower for the tested cigarette smoke fraction compared to a simple furans mixture, indicative for the presence of inhibitory effects in a complex mixture such as cigarette smoke
- The formation rate for furans reactive metabolites for lung subcellular fractions is significantly lower compared to liver, except for furan using S9 fraction
- Human liver and lung S9 fractions generate reactive sulfate conjugates for 5-methyl-2-furanmethanol and 5-hydroxymethylfurfural, with activity for lung S9 fraction more than 10 times lower compared to liver S9

References

- Peterson L. A. Reactive metabolites in the biotransformation of molecules containing a furan ring. Chem. Res. Toxicol. 26:6 (2013).
- Zhouhong Wang et al. Effect of cellulose crystallinity on the formation of a liquid intermediate and on product distribution during pyrolysis. Journal of Analytical and Applied Pyrolysis. 100: 56-66 (2013)
- Takakusa H. et al. Markers of Electrophilic Stress Caused by Chemically Reactive Metabolites in Human Hepatocytes. Drug Metabolism and Disposition. Vol. 36, No. 5 (2008).
- University of Kentucky (www.3r4f.com), (http://www2.ca.uky.edu/refcig/3R4F%20Preliminary%20Analysis.pdf)



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